Contents lists available at ScienceDirect



Palaeogeography, Palaeoclimatology, Palaeoecology

journal homepage: www.elsevier.com/locate/palaeo



Influence of dissolved oxygen on secular patterns of marine microbial carbonate abundance during the past 490 Myr



Robert Riding^{a,*}, Liyuan Liang^b, Jeong-Hyun Lee^c, Aurélien Virgone^d

^a Department of Earth and Planetary Sciences, University of Tennessee, Knoxville, TN 37996-1526, USA

^b Geoconsultants Corporation, Kevil, KY 42053, USA

^c Department of Geology and Earth Environmental Sciences, Chungnam National University, Daejeon 34134, Republic of Korea

^d TOTAL CSTJF, Avenue Larribau, F-64018 Pau Cedex, France

ARTICLE INFO

Keywords: Bacteria Calcification Diversity Invertebrate Oxygenation

Phanerozoic

ABSTRACT

Shallow marine benthic microbial carbonate sediments declined episodically during the Phanerozoic. They were relatively abundant during the Cambrian/mid-Ordovician, late Devonian/Mississippian, mid-Permian/mid-late Triassic, and to a lesser degree in the late Jurassic/early Cretaceous. They were least abundant when invertebrate, especially reef, diversity was relatively high in the mid-Ordovician/late Devonian, early-mid Permian, late Triassic, mid-late Jurassic, and early Cretaceous to present-day. These intervals of microbial carbonate abundance and invertebrate diversity generally alternate transitionally with one-another; but 'lows' and periods of overlap also occur. 'Lows', when benthic microbial carbonate abundance and invertebrate diversity both declined, are apparent during the Pennsylvanian and early-mid Jurassic. Overlaps, when microbial carbonates and invertebrates were both relatively common, occur in the early Cambrian and late Jurassic. Overall, these patterns broadly support suggestions that microbial carbonate abundance declined as calcified invertebrates increased. To further explore these relationships, we compared microbial carbonate abundance and invertebrate diversity with estimates of changes in sea-surface temperature and marine dissolved oxygen for the past 490 Myr. This analysis suggests that invertebrate diversity varied directly with oxygen availability. Invertebrate diversity was low when dissolved oxygen was low in the early Ordovician, late Devonian, early Triassic, and early-mid Jurassic. It appears that, by reducing invertebrate diversity, low oxygen levels favored microbial carbonates. At the same time, low oxygen levels likely stimulated anaerobic metabolisms favoring carbonate precipitation. This study suggests that dissolved oxygen was a major control on secular patterns of marine microbial carbonate abundance during the Phanerozoic, together with marine temperature and carbonate saturation state.

1. Introduction

Benthic microbial carbonates form by the calcification of bacterialalgal mats and biofilms (Burne and Moore, 1987; Dupraz et al., 2009) and have been common components of marine limestones and dolostones since the Archean (Peters et al., 2017). Their development was significantly affected by changes in marine life and environments during the Phanerozoic (the past 541 Myr). Microbial carbonates remained abundant and widespread in shallow seas during the Cambrian but subsequently declined and have been relatively scarce for the past 100 Myr (Fischer, 1965; Garrett, 1970). This decline was not linear; peaks of microbial carbonate abundance (~500, ~370, ~250, and ~150 Ma; Kiessling, 2002) alternate with intervals of increased invertebrate diversity and successively decrease in magnitude (Riding, 2005). The Phanerozoic decline of microbial carbonates is thought to mainly reflect the combined effects of reduced seawater carbonate saturation and increased invertebrate competition (Fischer, 1965; Garrett, 1970; Webb, 1996; Riding, 2005; Riding and Liang, 2005a; Yao et al., 2016). Seawater carbonate saturation state is mainly governed by calcium and carbonate ion activity (Plummer and Sundquist, 1982), but the factors that drive long-term changes in invertebrate abundance and diversity are more debatable. To further elucidate long-term controls on microbial carbonates, and particularly their relationships with invertebrates, we compared estimates of sea-surface temperature (SST) and dissolved oxygen in tropical seawater, with microbial carbonate abundance and benthic invertebrate diversity (as a proxy for competition) from the late Cambrian (~490 Ma) to the present-day (see Methods, below). This approach was stimulated by research that has

https://doi.org/10.1016/j.palaeo.2018.10.006

Received 13 August 2018; Received in revised form 8 October 2018; Accepted 8 October 2018 Available online 09 October 2018 0031-0182/ © 2018 Elsevier B.V. All rights reserved.

^{*} Corresponding author.

E-mail address: rriding@utk.edu (R. Riding).

linked the transition from microbial carbonate to skeletal carbonates during the early-mid Ordovician to changes in temperature and oxygenation (Berry and Wilde, 1978; Webby, 2002; Servais et al., 2008, 2010, 2016; Trotter et al., 2008; Saltzman et al., 2011; Kah et al., 2016; Rasmussen et al., 2016; Young et al., 2016; Edwards et al., 2017; Lee and Riding, 2018). A strong reciprocal relationship between oxygen and life (Nursall, 1959; Berkner and Marshall, 1964; Cloud, 1968; Holland, 1994; Berner et al., 2007; Knoll, 2014) has profoundly influenced the diversity of both invertebrates (Childress and Seibel, 1998; Levin, 2003; Seibel, 2011; Verberk et al., 2011; Ferguson et al., 2013; Wood and Erwin, 2018) and vertebrates (Berner et al., 2007). The rise of eukarvotes, particularly invertebrates, has been attributed to marine oxygenation (Nursall, 1959; Sperling et al., 2013; Knoll, 2014; Reinhard et al., 2016). Oxygen levels are thought to have risen significantly in the Neoproterozoic, ~800 Ma (Och and Shields-Zhou, 2012; Lenton et al., 2014; Lyons et al., 2014), but then more slowly during the early Paleozoic (Dahl et al., 2010; Lenton et al., 2016). Elevated ('greenhouse effect') temperatures during the mid-late Cambrian (Fischer, 1984; McKenzie et al., 2014) resulted in low levels of dissolved oxygen in seawater, and marine oxygenation probably did not recover until the mid-Ordovician (Edwards et al., 2017; Lee and Riding, 2018). Oxygen levels at or above those of the Cenozoic probably did not occur until the Silurian (Royer et al., 2014, their Fig. 2b) or Devonian (Wallace et al., 2017), and remained subject to periods of significant reduction until the Early Cretaceous (Berner, 2009; Lu et al., 2018).

Our comparisons indicate that during the past 490 Myr, invertebrate diversity was highest when dissolved oxygen levels were relatively high, whereas microbial carbonates were most abundant when dissolved oxygen levels were low, so long as carbonate saturation state was high. It appears that microbial carbonate abundance declined as invertebrate diversity increased with dissolved oxygen. In addition to competition for space, it is likely that invertebrate skeleton formation also reduced the CaCO₃ available for microbial carbonates. Our comparisons further suggest that intervals of microbial and invertebrate carbonate abundance over the past 490 Mya have not simply alternated between one another. In addition to transitions between intervals of microbial carbonate and invertebrate abundance, there are examples of overlap and also extended 'lows'. 'Lows', when microbial carbonate abundance and invertebrate diversity were both reduced, 323-299 and 201-170 Ma, may at least in part be due to lower temperatures. Examples of overlap, when invertebrates and microbial carbonates were both relatively abundant, occur ~525-510 and 164-152 Ma.

These long-term patterns suggest that invertebrates interacted competitively with microbial carbonates at several levels, and also responded to different physicochemical influences. Whereas oxygen promoted marine invertebrate diversity, microbial carbonate abundance increased when oxygen was low. In addition to reducing invertebrate diversity, low oxygen could have favored microbial carbonate formation by promoting anaerobic metabolisms. A further effect involves increased temperature, which tends to both raise carbonate saturation and decrease the level of dissolved oxygen. Thus, elevated temperature (e.g., in the late Cambrian and during the Permian–Triassic transition) would be expected to have inherently favored microbial carbonate formation, in addition to reducing invertebrate diversity.

We therefore propose that Phanerozoic patterns of microbial carbonate abundance and invertebrate diversity largely reflect the interplay of physicochemical factors. Microbial carbonate formation has tended to increase with carbonate saturation state, whereas invertebrate diversity has increased with the level of dissolved oxygen level. These results help to clarify the factors underlying the distinctive fluctuating pattern of abundance of microbial and invertebrate shallow marine carbonate sediments since the Cambrian. By influencing invertebrate diversity, dissolved oxygen has significantly influenced the abundance of marine microbial carbonates throughout the Phanerozoic.

2. Methods

We examined relationships among dissolved oxygen, sea-surface temperature, invertebrate genus richness (as a rough proxy for abundance/competition) and microbial carbonate abundance, over the past 490 Myr.

2.1. Microbial carbonate abundance

Stromatolite decline, measured by reduction in number of form taxa during the Neoproterozoic (Awramik, 1971) and reduction in overall abundance since the early Ordovician (Cloud and Semikhatov, 1969). has been linked to increased eukarvote competition and reduced calcification (Fischer, 1965, pp. 1208-1209). In addition to stromatolites (Peters et al., 2017, their Fig. 2), calcified cyanobacteria (Riding, 1992; Arp et al., 2001), reefal microbial carbonates (Kiessling, 2002, their Fig. 16) and microbialites in general (Myshrall et al., 2014, their Fig. 6.2) together, all exhibit a markedly episodic non-linear pattern of decline over the past 500 Myr. Microbial carbonates are locally abundant in the Silurian (e.g., Soja and Riding, 1993; Sheehan and Harris, 2004; Peters et al., 2017, their Fig. 2) and early Devonian (e.g., Matysik et al., 2015), with a minor peak ~420-410 Ma (Kiessling, 2002, his Fig. 16). However, the main peaks of microbial carbonate (MC) abundance (Kiessling, 2002, his Fig. 16; see Riding, 2005, 2006) recognized here are MC1: Cambrian/mid-Ordovician (541-465 Ma), MC2: latest Devonian/Mississippian (372-323 Ma), MC3: mid-Permian to mid-late Triassic (265-227 Ma) and MC4: late Jurassic-early Cretaceous (164-133 Ma). These peaks, respectively 76, 49, 38, and 31 Myr in duration and 93, 58, and 63 Myr apart, steadily diminish in magnitude over time (Fig. 1A). These data are currently the best available. Their improvement by future compilation of Phanerozoic microbial carbonate occurrences will doubtless refine and inform the conclusions reached here.

2.2. Invertebrate diversity

We compared these microbial carbonate abundance data (Kiessling, 2002, his Fig. 16; Riding and Liang, 2005b, their Fig. 1) (Fig. 1A), with published assessments of marine invertebrate diversity data (Mayhew et al., 2012, 'shareholder quorum subsampling' (SQS) values from supplementary data; Zaffos et al., 2017, their Fig. 2a) (for details of shareholder quorum subsampling see Alroy, 2010), and also with a compilation of bryozoan, cnidarian and poriferan diversity data (from http://fossilworks.org/ accessed 8th May 2018; see Supplemental Data) to represent invertebrate reef-builders (Fig. 1C). Throughout our discussion we use the terms 'richness' and 'diversity' interchangeably. Based on these data compilations we recognize the following main intervals of elevated invertebrate (I), particularly reefal, genus diversity: I1: 465-372 Ma (mid-Ordovician/late Devonian), I2: 299-265 Ma (early/mid Permian), I3: 227-201 Ma (late Triassic), I4: 170-152 Ma (mid/late Jurassic), I5: 133-0 Ma (early Cretaceous/present day) (Fig. 1). As with the microbial carbonate abundance data, future compilations that refine these intervals will modify the conclusions reached here.

2.3. Temperature and oxygenation

We used sea-surface temperatures (Mayhew et al., 2012; Royer et al., 2004), marine dissolved oxygen values calculated from atmospheric oxygen estimates of Royer et al. (2014), and seawater carbonate saturation state values calculated by Riding and Liang (2005b, their Fig. 1) (Fig. 1B). Numerical geological ages used throughout are those of the International Chronostratigraphic Chart (Cohen et al., 2013).

We calculated dissolved O2 in tropical surface seawater for the past



Fig. 1. (A) Microbial carbonate relative abundance in marine reefs, from Kiessling (2002, his Fig. 16), using Golonka and Kiessling's (2002, his Fig. 1, pp. 13-18) time intervals for supersequences (see Riding, 2005, his Fig. 1). Main peaks of abundance occur in the Cambrian-earliest Ordovician (541-465 Ma), latest Devonian-mid-Mississippian (372-323 Ma), mid-Permian to mid-late Triassic (265-227 Ma) and late Jurassic-early Cretaceous (164-133 Ma). These peaks, separated by intervals of lower abundance, steadily decline in magnitude throughout the Phanerozoic. Geological time-scale from Cohen et al. (2013). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

(B) Sea-surface temperature (Royer et al., 2004, their Fig. 4, their red curve, compiled by Mayhew et al., 2012, their supp. data), marine dissolved oxygen calculated from atmospheric oxygen estimates of Royer et al. (2014, their Fig. 2b), and seawater calcite saturation state calculated by Riding and Liang (2005b, their Fig. 1).

(C) Three estimates of Phanerozoic marine invertebrate genus richness. M: shareholder quorum subsampling (SQS) values from the supplementary data in Mayhew et al. (2012). Z: Range-through marine invertebrate generic richness calculated in million-year increments (Zaffos et al., 2017, their Fig. 2a); R: compilation of bryozoan, cnidarian and poriferan genus richness data to represent invertebrate reef-builders. This latter compilation includes all fossilized members of these phyla, mainly reef-

building organisms. Porifera include archaeocyaths, hexactinellids, lithistids, sclerosponges, sphinctozoans and stromatoporoids. Cnidaria include rugose, tabulate, and scleractinian corals. Bryozoa include gymnolaemates and stenolaemates.

490 Myr (latest Cambrian to present-day) (Fig. 1B), using modeled atmospheric oxygen estimates from Royer et al. (2014, their Fig. 2b) and sea surface temperature anomaly data from Royer et al. (2004, see their Fig. 4, red curve), compiled by Mayhew et al. (2012, 'temperature' column in their supp. dataset SD01x.xlsx). Dissolved oxygen in surface seawater depends on atmospheric oxygen composition together with water temperature, pressure, and salinity. We made the following assumptions: present-day tropical sea-surface temperate 25 °C, water depth of 0 m (i.e., surface of seawater at 1 atm), and salinity 35 parts per thousand. According to Henry's law, at constant temperature and pressure, the amount of O2 gas that dissolves in water is directly proportional to the partial pressure of O2 gas in equilibrium with water. In seawater, the presence of salt lowers the dissolved O2. A table of solubility of oxygen in seawater at ${\sim}35^0/_{00}$ salinity and 20% partial pressure of oxygen gas is given by the Engineering ToolBox (https:// www.engineeringtoolbox.com/oxygen-solubility-water-d_841.html) (Table S1, Supplemental Data).

To obtain the Henry's law constants for surface seawater any given temperature we derived a polynomial function as follows. Using the tabulated dissolved oxygen data (Table S1, Supplemental Data), we first calculated discrete K_H values corresponding to the given temperatures (every 5 °C) at 20% partial pressure of O_2 (Table S2, Supplemental Data). We then obtained a polynomial function by fitting the K_H versus temperature curve (Fig. S1, Supplemental Data):

$$K_{\rm H} = -0.0047 T^3 + 0.68 T^2 - 42.6 T + 1741 \tag{1}$$

where T is temperature in °C.

We used the above polynomial function and calculated the predicted K_H at a given temperature using sea-surface temperature values from Royer et al. (2004), compiled by Mayhew et al. (2012) with a baseline temperature of 25 °C.

Next, we used Henry's law equation to calculate the dissolved oxygen in μ M (Table S3, Supplemental Data):

$$C = K_{\rm H} \times p_{\rm O2} \tag{2}$$

where the partial pressure of oxygen, p_{O2} is from Royer et al. (2014, their Fig. 2b). The computed dissolved oxygen values are displayed in Fig. 1B.

3. Results

3.1. Invertebrate diversity and dissolved oxygen

Oxygen levels broadly track our measures of invertebrate diversity for the past 490 Myr (Fig. 1B). Our comparison of Mayhew et al.'s (2012) diversity data (using *raw* values of 'shareholder quorum subsampling', SQS) and dissolved oxygen shows positive correlation: r = +0.384, $p(\alpha) = 0.01 < 0.05$, n = 44 (Table S4, Supplemental Data). Invertebrate diversity is generally high when dissolved oxygen is high, although a conspicuous exception is when oxygen was high and diversity low ~300 Ma. This could reflect the effect on diversity of Ice Age conditions at that time, such as low temperature and low sea-level. In contrast, diversity was generally low when oxygen was low, e.g., ~475, 370, 250, 180 Ma, likely reflecting the effect of increased tendency to seasonal and/or localized hypoxia. Based on our calculated values, dissolved oxygen levels near or below 150 µmol per liter (µM) coincide with low invertebrate diversity in the Floian–Dapingian (479–468 Ma), early Famennian (369 Ma) and early Jurassic (193–171 Ma) (Fig. 1).

3.2. Invertebrate diversity and temperature

Using standardized subsampling techniques Mayhew et al. (2012, p. 15141, and their Fig. 1D), reported positive correlation between diversity (SQS) and temperature, r = +0.289. However, using Mayhew et al. (2012)'s *raw* SQS values we find diversity and temperature to be poorly correlated: r = -0.10, $p(\alpha) > 0.05$, n = 44 (Table S4, Supplementary Data). Our comparisons suggest that invertebrate diversity and sea-surface temperature can correspond (e.g., both relatively high ~400 and 190 Ma; both relatively low ~300 Ma), and also diverge (e.g., ~500, 260, 20 Ma) (Fig. 1).

3.3. Microbial carbonates and dissolved oxygen

Broadly, peaks of microbial carbonate abundance (e.g., \sim 500, 370, 237, 150 Ma) occur when dissolved oxygen values were at or below \sim 170 µM (Fig. 1). This appears to reflect the effect of reduced invertebrate diversity (and therefore competition), in response to low oxygen levels, on microbial carbonate formation.

3.4. Microbial carbonates, temperature and carbonate saturation state

Microbial carbonate abundance is generally positively correlated with sea-surface temperature, which tends to raise carbonate saturation state, and intervals of microbial carbonate abundance also broadly correspond with increased levels of calculated carbonate saturation state (Fig. 1). However, extended periods occur when microbial carbonates were not very abundant even though calculated carbonate saturation was relatively high; notably the mid-Ordovician to mid-Devonian and mid-late Cretaceous. These discrepancies likely reflect increased biocontrolled calcification during these intervals, which was not taken into consideration in the saturation state values calculated by Riding and Liang (2005a, their Fig. 5) (see Section 4.5).

3.5. Invertebrate diversity and microbial carbonates

Broadly, peaks of microbial carbonate abundance correspond with low invertebrate diversity, and peaks of invertebrate diversity ~465–372, 299–265, 133–0 Ma correspond with low microbial carbonate abundance (Fig. 1). Conversely, there are at least two intervals when invertebrate diversity and microbial carbonate abundance were both low: ~323–299, 201–170 Ma. These are times when microbial carbonate abundance appears to have been depressed by low carbonate saturation state (Riding and Liang, 2005a, 2005b) and when low temperatures (together with low dissolved oxygen levels, ~201–170 Ma) can be expected to have lowered invertebrate diversity. Overall, therefore, on the scales of these data, microbial carbonates were more abundant when invertebrate diversity was low, except when temperature and/or carbonate saturation state were also low.

3.6. Transitions, 'lows', overlaps

Our data support the observation that intervals of microbial and invertebrate carbonate abundance have broadly alternated with one another over long time scales during the Phanerozoic (Riding, 2005). In addition, our more precise delimitation of variations in invertebrate diversity also indicate three types of change between these intervals: transitions, 'lows', overlaps. Well-defined transitions occur between intervals of microbial carbonate and invertebrate abundance in the mid-Ordovician, late Devonian, late Permian, and mid-Triassic. An early Cretaceous transition is less well-defined due to the lower abundance of microbial carbonates at that time. Extended 'lows', when microbial carbonate abundance and invertebrate diversity were both reduced 323-299 and 201-170 Ma may, at least in part, be due to low temperatures. Overlaps, when invertebrates and microbial carbonates were both relatively abundant, occur ~525-510 and 164-152 Ma. Both of these examples involve sponges and microbial carbonates: mainly archaeocyaths in the Cambrian and mainly hexactinellids and lithistids in the late Jurassic. This may reflect the ability of sponges to tolerate conditions that also favored microbial carbonate formation (Lee and Riding, 2018). Further studies of microbial-invertebrate transitions are likely to reveal additional details and complexities.

3.7. Summary of intervals

Four intervals when microbial carbonate (MC) abundance was increased and invertebrate diversity was reduced occur during the Phanerozoic: MC1: Cambrian/mid-Ordovician (541-465 Ma), MC2: latest Devonian/Mississippian (372-323 Ma), MC3: mid-Permian/midlate Triassic (265-227 Ma), and MC4: late Jurassic/early Cretaceous (164-133 Ma) (Fig. 1). These MC intervals alternate - often transitionally - with intervals of relatively high invertebrate (I), especially reef, genus diversity when microbial carbonates were less common (Fig. 2). The main intervals of increased invertebrate diversity are I1: mid-Ordovician/late Devonian (465-372 Ma), I2: early/mid Permian (299-265 Ma), I3: late Triassic (227-201 Ma), I4: mid/late Jurassic (170–152 Ma), I5: early Cretaceous/present day (133–0) Ma (Fig. 2). In addition to transitions, there are lows' between some of these intervals when microbial carbonates and invertebrates were both relatively reduced, e.g., during the Pennsylvanian (323-299 Ma) and early Jurassic (201-170 Ma). There are also at least two periods of overlap when invertebrates and microbial carbonates were both relatively abundant: early-mid Cambrian (~525-510 Ma), within MC1, when archaeocyaths were common, and late Jurassic (164-152 Ma), between I4 and MC4, also when sponges were common (Fig. 2).

4. Discussion

4.1. Invertebrate diversity and dissolved oxygen

Our comparisons suggest that marine invertebrate diversity has broadly tracked marine oxygenation since the Cambrian. Diversity was low when dissolved oxygen levels were low (and SST high), ~475, 370, 250, 180 Ma (Fig. 1). In addition, our comparison with Mayhew et al.'s (2012) raw shareholder quorum subsampling (SQS) data shows that diversity and oxygen are significantly correlated: r = +0.384 (p $(\alpha) < 0.05, n = 44$; see Section 3.1). Marine oxygenation has been linked to late Proterozoic and Cambrian animal evolution (see Introduction, above) and it has been proposed that 'oxygen was the chief limiting factor on the density of animal populations' in the Early Paleozoic (Fischer, 1965, p. 1211). However, whereas oxygen levels have been invoked to interpret terrestrial evolution (Berner, 2006), patterns of marine invertebrate diversity during the Phanerozoic as a whole have more often been linked to factors such as continental flooding, nutrient supply, temperature, tectonics, seafloor spreading rate, and global biogeography (e.g., Martin, 1996; Mayhew et al., 2008; Benton, 2009; Cárdenas and Harries, 2010; Hannisdal and Peters, 2011; Cermeño et al., 2017), as well as to 'intrinsic traits of animal taxa' (Stanley, 2007).

Despite relatively high present-day levels of atmospheric oxygen (Berner, 2009), zones of low oxygenation associated with upwelling



Fig. 2. Phanerozoic Intervals characterized by microbial and invertebrate skeletal carbonate. Geological time-scale from Cohen et al. (2013). There are four intervals of increased microbial carbonate abundance: MC1: 541-465, MC2: 372-323, MC3: 265-227, and MC4: 164-133 Ma. These alternate with intervals of relatively high invertebrate, especially reef, diversity when microbial carbonates were less common: I1: 465-372, I2: 299-265, I3: 227-201, I4: 170-152, I5: 133-0 Ma. 'Lows', when microbial carbonates and invertebrates were both relatively reduced, occur 323-299 and 201-170 Ma. Two intervals of overlap, when invertebrates and microbial carbonates were both relatively abundant, occur within MC1, ~525-510 Ma, and between I4 and MC4, 164-152 Ma.

and stratification are locally and seasonally common in restricted seas, especially where there is nutrient influx, and develop seasonally as oxygen-minimum zones (OMZs) in the open ocean (Stramma et al., 2008; Levin, 2018). Some organisms, such as many sponges, tolerate low oxygen. Present-day hexactinellids, for example, tolerate hypoxia (< 1.4 mL/L) (Chu and Tunnicliffe, 2015) and can survive anoxia (oxygen concentrations < 0.2 mL/L) (Tunnicliffe, 1981). However, many invertebrates are stressed by these conditions (Levin et al., 2009; Rabalais et al., 2010; Stramma et al., 2010), and even water conventionally regarded as only marginally hypoxic (2 mg O2/L) may be lethal or sublethal for many species (Vaquer-Sunyer and Duarte, 2008). In present-day reefs, the coral Acropora yongei can be significantly harmed by low oxygen (2-4 mg/L) (Haas et al., 2014), and coral species richness is reduced below 2 mg/L (Altieri et al., 2017, their Fig. 2). In addition to absolute levels of oxygen, short-term variability in oxygen concentration can also be severely limiting (Matabos et al., 2012). Several mass extinction events have been specifically linked to low dissolved oxygen, e.g., Frasnian-Famennian, Permian-Triassic, and end-Triassic (Meyer and Kump, 2008; van de van de Schootbrugge and Wignall, 2015; Jost et al., 2017; Levin, 2018, their supp. Fig. 3). Over time, it is likely that variations in dissolved oxygen have strongly influenced marine sessile invertebrate reef-builders, and intervals of anoxia broadly correspond with increased abundance of microbial reefs (Knoll and Fischer, 2011, their Fig. 4.1). Under the low atmospheric values calculated for some past intervals, particularly when overall temperatures were increased, similar low-oxygen conditions and their effects, are likely to have been expanded in time and space, e.g., late Cambrian-early Ordovician, late Devonian, Early Triassic, and Jurassic-Early Cretaceous (Fig. 1).

Exceptionally preserved biotas (Allison and Briggs, 1993) and oceanic anoxic events (OAEs) (Jenkyns, 2010) also tend to occur within extended intervals with low levels of dissolved O₂. Well-known examples include mid–late Cambrian and early Ordovician Burgess Shale type faunas (Butterfield, 1995; Van Roy et al., 2010; Lerosey-Aubril et al., 2017), Late Devonian Kellwasser events (Bond et al., 2004), Mississippian Shrimp Beds (Aldridge et al., 1993), the late Permian mass extinction (Brennecka et al., 2011; Lau et al., 2016), and the early Toarcian event (Posidonienschiefer, T-OAE, ~183 Ma) (Jenkyns, 2010).

We propose that oxygenation exerted a significant influence on marine invertebrates; specifically, that low levels of dissolved oxygen limited diversity. This is consistent with recognition that present-day low oxygen (hypoxia) can reduce marine invertebrate diversity (e.g., Vaquer-Sunyer and Duarte, 2008; Levin et al., 2009; Rabalais et al., 2010; Stramma et al., 2010). In addition to its direct effect, oxygenation

can increase availability of key nutrients such as phosphorus (Laakso and Schrag, 2017). The Cambrian-Ordovician transition from microbial to invertebrate carbonates (Adachi et al., 2009) exemplifies these inferred effects. In the late Cambrian and early Ordovician, microbial carbonates were abundant whereas invertebrate diversity appears to have been limited by low levels of dissolved oxygen that reflect the combined effects of elevated marine temperatures (Trotter et al., 2008) and relatively low atmospheric oxygen levels (Webby, 2002; Edwards et al., 2017, their Fig. 2). During the early-mid Ordovician, microbial carbonate abundance declined and invertebrates diversified, as temperature declined and dissolved O₂ is estimated to have increased from 100 to 150 µM (Lee and Riding, 2018). These changes were the prelude to a protracted phase that lasted until the beginning of the late Devonian, during which – very broadly – invertebrate diversity tended to be high and microbial carbonate abundance low. Subsequent episodes of increased microbial carbonate abundance (~372-323, 265-227, 164-133 Ma) coincided with low levels of dissolved oxygen and low invertebrate diversity (Fig. 1). We therefore suggest that the distinctive oscillating pattern of microbial carbonate abundance and invertebrate diversity that characterizes much of the Phanerozoic was underpinned by changes in marine oxygenation that significantly affected invertebrates and, thereby, microbial carbonates. Provided carbonate saturation was elevated, microbial carbonate abundance increased when low dissolved oxygen levels limited invertebrate diversity.

4.2. Invertebrate diversity and temperature

Links between invertebrate diversity and temperature (Fischer, 1960) continue to be debated (Currie, 1991; Clarke and Gaston, 2006; Mittelbach et al., 2007; Kiessling, 2009; Gillman and Wright, 2014). Using standardized subsampling techniques, Mayhew et al. (2012, p. 15141, and their Fig. 1D), found positive correlation between invertebrate diversity (SQS) and seawater temperature, r = +0.289, for the Phanerozoic as a whole. However, the same diversity and temperature data show times of both broadly positive correlation (e.g., ~400, 300, 190 Ma) and negative correlation (e.g., 500, 260, 20 Ma), and using Mayhew et al. (2012) raw SQS values, we calculate overall slightly negative correlation, r = -0.10 (p(α) > 0.05, n = 44; see Section 3.2), between diversity and temperature. Two conspicuous divergences that occur between diversity and temperature, ~500 and 260 Ma, may respectively reflect diversity decline as elevated temperatures depressed dissolved oxygen levels. Conversely, increased diversity during the great Ordovician biodiversification event (GOBE) can be linked to a combination of temperature decline and increased oxygenation (Webby, 2002; Trotter et al., 2008; Edwards et al., 2017, their Fig. 3; Lee and Riding, 2018, their Fig. 7).

4.3. Microbial carbonates and dissolved oxygen

These links between dissolved oxygen and invertebrate diversity affected microbial carbonate abundance (Fig. 1). Our comparisons suggest that microbial carbonate abundance increased when invertebrate competition was reduced by low dissolved O2 levels. In addition, many microbes not only tolerate but are well-adapted to low O₂. Thus, correspondence between microbial carbonate abundance and low levels of dissolved oxygen could reflect increase in anaerobic bacterial metabolisms that can promote carbonate precipitation (Grotzinger and Knoll, 1995; Higgins et al., 2009; Schrag et al., 2013; Sun and Turchyn, 2014), such as sulfate reduction. Overall, Phanerozoic microbial carbonates appear to have responded positively to a combination of low dissolved O2, elevated temperature, and increased carbonate saturation state, and to absence of invertebrates, just as they seem to have done in the Precambrian. As the Phanerozoic progressed, these conditions became scarcer. Present-day normal marine carbonates are overwhelmingly dominated by the skeletal remains of algae and invertebrates (Leeder, 1982; Schneider et al., 2000), whereas benthic microbial carbonates are much more restricted. Microbial crusts in Quaternary reefs exemplify an anaerobic marine heavily calcified stromatolite 'refuge', now declining as natural acidification lowers carbonate saturation state (Riding et al., 2014).

4.4. Invertebrate diversity and microbial carbonate abundance

The inverse relationship between microbial carbonate abundance and invertebrates based on Sepkoski Jr.'s (1997) data (Riding, 2005, his Fig. 2; Riding and Liang, 2005a; Riding, 2006), is supported here by comparisons with recent compilations of invertebrate diversity (Mayhew et al., 2012, SQS from their supplementary data; Zaffos et al., 2017), as well our compilation of reef invertebrate diversity data (Fig. 1). Overall, intervals characterized by increased microbial carbonate abundance correspond with lower invertebrate diversity: Cambrian-earliest Ordovician (541-480 Ma), latest Devonian-Mississippian (372-323 Ma), mid-Permian-late Triassic (265-227 Ma), and late Jurassic-earliest Cretaceous (164-133 Ma). Despite lack of clear correlation between all mass extinctions and increased microbial carbonate abundance (Riding, 2005), this pattern is consistent with the long-held view that invertebrate competition has significantly limited microbial carbonate abundance (Fischer, 1965; Garrett, 1970; Awramik, 1971). Fischer (1965) suggested that, in addition to competition, stromatolite may have declined since the Ordovician because they 'became less mineralized'. Microbial carbonate abundance broadly correlates positively with calculated seawater carbonate saturation state (Riding and Liang, 2005a).

The likelihood that microbial carbonate abundance reflects a response to both competition and carbonate saturation state raises the question of the relative importance of these influences (Riding, 2005). Microbial carbonates were not always abundant when invertebrate diversity was low (e.g., ~300 and 200 Ma), and this can tentatively be attributed to low carbonate saturation state and/or low temperatures. Conversely, microbial carbonates were not always abundant when carbonate saturation state was high, for example the Silurian to late Devonian. Since this was an interval of generally increased invertebrate diversity, it seems reasonable to conclude that increased benthic invertebrate competition limited microbial carbonates by competing for both space and calcium carbonate, similar to the effect on seawater carbonate saturation of the Mesozoic-Cenozoic rise of calcified plankton (Sandberg, 1975; Wilkinson and Walker, 1989; Riding, 1993; Webb, 1996; Riding and Liang, 2005a).

4.5. The nature of competition

We used measures of diversity through time to represent invertebrate abundance and thereby competition with microbial carbonates. Competition for space and resources is often intense among benthic, especially reef, communities (McCook et al., 2001; Taylor and Wilson, 2003; Chadwick and Morrow, 2011; Svensson and Marshall, 2015). Organisms exhibit overgrowth, grazing and predation (Taylor and Wilson, 2003) and compete for limiting resources such as food, space and oxygen (Ferguson et al., 2013). Invertebrate competition with microbial carbonates has been suggested to include grazing and burrowing (Garrett, 1970), boring (Myshrall et al., 2014), substrate competition (Webb, 2005), and the effects of bioclastic and pelleted sediment production (Pratt, 1982). In addition, there could be competition not only for space but also for CaCO₃ supply. Carbonate saturation state values calculated by Riding and Liang (2005a) do not take account of biocontrolled CaCO3 removal. Consequently, elevated saturation state levels calculated for the Ordovician to Devonian do not preclude the likelihood that increase in invertebrate biocalcification during that extended interval would have lowered seawater carbonate saturation sufficiently to limit microbial calcification. Riding (2005, p. 33) observed that 'metazoan interference in microbial carbonate formation could include competition for the CaCO₃ required for microbial calcification'.

In addition, microbes can compete with metazoans (Webb, 2005). At the present-day, for example, cyanobacterial mats can inhibit coral recruitment and promote reef degradation (Ford et al., 2018). During the Cambrian, ability to exploit low-oxygen conditions may have enabled microbial carbonates to outcompete metazoan reef builders such as sponges and corals, whereas progressive Ordovician marine oxygenation subsequently promoted diversification of skeletal metazoans that could outcompete microbial carbonates for seafloor space (Lee and Riding, 2018). Reef competition is complex due to the variety of substrates and seafloor morphology, such as oxygen-poor crypts, sedimentfilled crevices, current-swept surfaces and exposed hardgrounds. Consequently, even during protracted intervals when microbial carbonates appear to have been relatively scarce, such as much of the Devonian (~410-372 Ma), reefal microbial carbonates were locally abundant and thrived as cryptic crusts in reefs (Pratt, 1982, 1995; Soja and Riding, 1993; Soja, 1994; de Freitas and Mayr, 1995; Webb, 1996; de Freitas and Nowlan, 1998; Sheehan and Harris, 2004; Matysik et al., 2015; Peters et al., 2017, their Fig. 2).

If microbial carbonates decline as competition increases (Garrett, 1970; Awramik, 1971), then they could increase when invertebrates decline. Thus, Schubert and Bottjer (1992, 1995) suggested that Early Triassic stromatolites represent a temporary resurgence in the aftermath of the end-Permian mass extinction. However, microbial carbonates only appear to increase after some (e.g., Late Devonian, Permian-Triassic) rather than all (e.g., end-Ordovician, end-Triassic (but see Ibarra et al., 2016), end-Cretaceous) mass extinction events, indicating that factors in addition to competition influence microbial carbonate development. This implicates carbonate saturation state as the key control since, in mass extinction aftermaths, microbial carbonate abundance increased at times when saturation state was high (e.g., Late Devonian and end-Permian), but not when it was low (e.g., Late Cretaceous) (Riding, 2005, 2006). Based on comparison of metazoan diversity data (Sepkoski Jr., 1997) and calculated seawater carbonate saturation state (Riding and Liang, 2005a), Riding and Liang (2005b) suggested that a combination of changes in carbonate saturation state and metazoan competition could account for Phanerozoic episodes of microbial carbonate (Kiessling, 2002, his Fig. 16) and calcified

cyanobacteria (Arp et al., 2001) abundance, which 'broadly coincide with intervals when carbonate saturation state was elevated and metazoan diversity reduced.' This supports Fischer's (1965) earlier inference. Thus, on the scales shown by these data, microbial carbonates were more abundant when invertebrate diversity was low, so long as carbonate saturation state was elevated.

5. Conclusions

Intervals of microbial carbonate abundance and reduced invertebrate diversity correspond with increased carbonate saturation state, elevated temperature, and low levels of dissolved oxygen (Fig. 1). Conversely, increased invertebrate diversity and lower microbial carbonate abundance occurred during intervals with lower temperatures and higher levels of dissolved oxygen. These secular patterns support previous inferences that, after the Cambrian, microbial carbonate abundance was limited by invertebrate competition and low seawater carbonate saturation state. In addition these patterns suggest a positive long-term link between invertebrate diversity and marine oxygenation. Low invertebrate diversity and increased microbial carbonate abundance correspond with low levels of dissolved oxygen, and invertebrate diversity increased and microbial carbonate abundance declined when dissolved oxygen levels increased. Significant influence of dissolved oxygen on invertebrate diversity, if confirmed, provides a valuable new insight into the development of Phanerozoic marine biotas.

Phanerozoic environmental factors that favored microbial carbonate formation are reminiscent of conditions inferred for much of the Archean and early-mid Proterozoic: elevated seawater saturation state, low O2 and elevated temperature, as well as absence of invertebrate competition. During the past ~500 Myr, decline in microbial carbonate abundance appears to have reflected global long-term reduction in carbonate seawater saturation state and temperature, as atmospheric CO₂ declined, together with increase in marine oxygenation that favored invertebrate diversification and competition. During the past ~100 Myr, oxygen and invertebrate diversity increased, temperatures declined, and marine microbial carbonates became relatively scarce. These patterns underscore the importance of seawater saturation state and invertebrate competition, which previous research has recognized as significant influences on microbial carbonate abundance. Additionally, they identify dissolved oxygen as a key factor influencing invertebrate diversification. This contributes to fundamental understanding of the physicochemical factors underlying the alternating pattern of microbial and invertebrate carbonate intervals during the Phanerozoic.

Acknowledgments

J-HL's research was supported by the National Research Foundation of Korea (grant 2016R1C1B1012104). RR's research was supported by TOTAL S.A. AV thanks Emmanuelle Vennin for stimulating discussion. We are grateful to Tom Algeo for excellent editorial support and guidance, and to Steve Kershaw, Gregg Webb and an anonymous reviewer for very helpful suggestions that helped to improve the final manuscript.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.palaeo.2018.10.006.

References

- Adachi, N., Ezaki, Y., Liu, J., Cao, J., 2009. Early Ordovician reef construction in Anhui Province, South China: a geobiological transition from microbial- to metazoandominant reefs. Sediment. Geol. 220, 1–11.
- Aldridge, R.J., Briggs, D.E.G., Smith, M.P., Clarkson, E.N.K., Clark, N.D.L., 1993. The

- anatomy of conodonts. Philos. Trans. R. Soc. Lond. Ser. B Biol. Sci. 340, 406–421. Allison, P.A., Briggs, D.E.G., 1993. Exceptional fossil record: distribution of soft-tissue preservation through the Phanerozoic. Geology 21, 527–530.
- Alroy, J., 2010. Fair sampling of taxonomic richness and unbiased estimation of origination and extinction rates. In: Alroy, J., Hunt, G. (Eds.), Quantitative Methods in Paleobiology. Vol. 16. pp. 55–80 The Paleontological Society Papers.
- Altieri, A.H., Harrison, S.B., Seemann, J., Collin, R., Diaz, R.J., Knowlton, N., 2017. Tropical dead zones and mass mortalities on coral reefs. Proc. Natl. Acad. Sci. 114, 3660–3665.
- Arp, G., Reimer, A., Reitner, J., 2001. Photosynthesis-induced biofilm calcification and calcium concentrations in Phanerozoic oceans. Science 292, 1701–1704.
- Awramik, S.M., 1971. Precambrian columnar stromatolite diversity: reflection of metazoan appearance. Science 174, 825–827.
- Benton, M.J., 2009. The Red Queen and the Court Jester: species diversity and the role of biotic and abiotic factors through time. Science 323, 728–732.
- Berkner, L.V., Marshall, L.C., 1964. The history of oxygenic concentration in the earth's atmosphere. Discuss. Faraday Soc. 37, 122–141.
- Berner, R.A., 2006. GEOCARBSULF: A combined model for Phanerozoic atmospheric O₂ and CO₂. Geochim. Cosmochim. Acta 70, 5653–5664.
- Berner, R.A., 2009. Phanerozoic atmospheric oxygen: new results using the
- GEOCARBSULF model. Am. J. Sci. 309, 603-606.
- Berner, R.A., Vandenbrooks, J.M., Ward, P.D., 2007. Oxygen and evolution. Science 316, 557–558.
- Berry, W.B.N., Wilde, P., 1978. Progressive ventilation of the oceans-an explanation for the distribution of the lower Paleozoic black shales. Am. J. Sci. 278, 257–275.
- Bond, D., Wignall, P.B., Racki, G., 2004. Extent and duration of marine anoxia during the Frasnian–Famennian (Late Devonian) mass extinction in Poland, Germany, Austria and France. Geol. Mag. 141, 173–193.
- Brennecka, G.A., Herrmann, A.D., Algeo, T.J., Anbar, A.D., 2011. Rapid expansion of oceanic anoxia immediately before the end-Permian mass extinction. Proc. Natl. Acad. Sci. 108, 17631–17634.
- Burne, R.V., Moore, L.S., 1987. Microbialites: organosedimentary deposits of benthic microbial communities. PALAIOS 2, 241–254.
- Butterfield, N.J., 1995. Secular distribution of Burgess-Shale-type preservation. Lethaia 28, 1–13.
- Cárdenas, A.L., Harries, P.J., 2010. Effect of nutrient availability on marine origination rates throughout the Phanerozoic eon. Nat. Geosci. 3, 430–434.
- Cermeño, P., Benton, M.J., Paz, O., Vérard, C., 2017. Trophic and tectonic limits to the global increase of marine invertebrate diversity. Sci. Rep. 7, 15969.
- Chadwick, N.E., Morrow, K.M., 2011. Competition among sessile organisms on coral reefs. In: Dubinsky, Z., Stambler, N. (Eds.), Coral Reefs: An Ecosystem in Transition. Springer, Dordrecht, pp. 347–371.
- Childress, J.J., Seibel, B.A., 1998. Life at stable low oxygen levels: adaptations of animals to oceanic oxygen minimum layers. J. Exp. Biol. 201, 1223–1232.
- Chu, J.W., Tunnicliffe, V., 2015. Oxygen limitations on marine animal distributions and the collapse of epibenthic community structure during shoaling hypoxia. Glob. Chang. Biol. 21, 2989–3004.
- Clarke, A., Gaston, K.J., 2006. Climate, energy and diversity. Proc. R. Soc. Lond. B Biol. Sci. 273, 2257–2266.
- Cloud, P.E., 1968. Atmospheric and hydrospheric evolution on the primitive earth. Both secular accretion and biological and geochemical processes have affected earth's volatile envelope. Science 160, 729–736.
- Cloud, P.E., Semikhatov, M.A., 1969. Proterozoic stromatolite zonation. Am. J. Sci. 267, 1017–1061.
- Cohen, K.M., Finney, S.C., Gibbard, P.L., Fan, J.-X., 2013. The ICS International
- Chronostratigraphic Chart. Episodes 36, 199–204.
 Currie, D.J., 1991. Energy and large-scale patterns of animal and plant species richness.
 Am. Nat. 137, 27–49.
- Dahl, T.W., Hammarlund, E.U., Anbar, A.D., Bond, D.P., Gill, B.C., Gordon, G.W., Knoll, A.H., Nielsen, A.T., Schovsbo, N.H., Canfield, D.E., 2010. Devonian rise in atmospheric oxygen correlated to the radiations of terrestrial plants and large predatory fish. Proc. Natl. Acad. Sci. 107, 17911–17915.
- de Freitas, T., Mayr, U., 1995. Kilometre-scale microbial buildups in a rimmed carbonate platform succession, Arctic Canada: new insight on Lower Ordovician reef facies. Bull. Can. Petrol. Geol. 43, 407–432.
- de Freitas, T.A., Nowlan, G.S., 1998. A new, major Silurian reef tract and overview of regional Silurian reef development, Canadian Arctic and north Greenland. Bull. Can. Petrol. Geol. 46, 327–349.
- Dupraz, C., Reid, R.P., Braissant, O., Decho, A.W., Norman, R.S., Visscher, P.T., 2009. Processes of carbonate precipitation in modern microbial mats. Earth Sci. Rev. 96, 141–162.
- Edwards, C.T., Saltzman, M.R., Royer, D.L., Fike, D.A., 2017. Oxygenation as a driver of the great Ordovician biodiversification event. Nat. Geosci. 10, 925–929.
- Ferguson, N., White, C.R., Marshall, D.J., 2013. Competition in benthic marine invertebrates: the unrecognized role of exploitative competition for oxygen. Ecology 94, 126–135.
- Fischer, A.G., 1960. Latitudinal variations in organic diversity. Evolution 14, 64-81.
- Fischer, A.G., 1965. Fossils, early life, and atmospheric history. Proc. Natl. Acad. Sci. 53, 1205–1215.
- Fischer, A.G., 1984. The two Phanerozoic supercycles. In: Berggren, W.A., Van Couvering, J.A. (Eds.), Catastrophes and Earth History: The New Uniformitarianism. Princeton University Press, Princeton, NJ, pp. 129–150.
- Ford, A.K., Bejarano, S., Nugues, M.M., Visser, P.M., Albert, S., Ferse, S.C.A., 2018. Reefs under siege—the rise, putative drivers, and consequences of benthic cyanobacterial mats. Front. Mar. Sci. 5, 18. https://doi.org/10.3389/fmars.2018.00018.
- Garrett, P., 1970. Phanerozoic stromatolites: noncompetitive ecologic restriction by

grazing and burrowing animals. Science 169, 171-173.

Gillman, L.N., Wright, S.D., 2014. Species richness and evolutionary speed: the influence of temperature, water and area. J. Biogeogr. 41, 39–51.

- Golonka, J., Kiessling, W., 2002. Phanerozoic time scale and definition of time slices. In: Kiessling, W., Flügel, E., Golonka, J. (Eds.), Phanerozoic Reef Patterns. SEPM Special Publication 72. SEPM, Tulsa, pp. 11–20.
- Grotzinger, J.P., Knoll, A.H., 1995. Anomalous carbonate precipitates: is the Precambrian the key to the Permian? PALAIOS 10, 578–596.
- Haas, A.F., Smith, J.E., Thompson, M., Deheyn, D.D., 2014. Effects of reduced dissolved oxygen concentrations on physiology and fluorescence of hermatypic corals and benthic algae. PeerJ e235, 2.
- Hannisdal, B., Peters, S.E., 2011. Phanerozoic Earth system evolution and marine biodiversity. Science 334, 1121–1124.
- Higgins, J.A., Fischer, W.W., Schrag, D.P., 2009. Oxygenation of the ocean and sediments: consequences for the seafloor carbonate factory. Earth Planet. Sci. Lett. 284, 25–33.
- Holland, H.D., 1994. Early Proterozoic atmospheric change. In: Bengtson, S. (Ed.), Early Life on Earth. Columbia University Press, New York, pp. 237–244.
- Ibarra, Y., Corsetti, F.A., Greene, S.E., Bottjer, D.J., 2016. A microbial carbonate response in synchrony with the end-Triassic mass extinction across the SW UK. Sci. Rep. 6, 19808.
- Jenkyns, H.C., 2010. Geochemistry of oceanic anoxic events. Geochem. Geophys. Geosyst. 11, Q03004.
- Jost, A.B., Bachan, A., van de Schootbrugge, B., Lau, K.V., Weaver, K.L., Maher, K., Payne, J.L., 2017. Uranium isotope evidence for an expansion of marine anoxia during the end-Triassic extinction. Geochem. Geophys. Geosyst. 18, 3093–3108. https://doi. org/10.1002/2017GC006941.
- Kah, L.C., Thompson, C.K., Henderson, M.A., Zhan, R., 2016. Behavior of marine sulfur in the Ordovician. Palaeogeogr. Palaeoclimatol. Palaeoecol. 458, 133–153.
- Kiessling, W., 2002. Secular variations in the Phanerozoic reef ecosystem. In: Kiessling, W., Flügel, E., Golonka, J. (Eds.), Phanerozoic Reef Patterns. SEPM Special Publication 72. SEPM, Tulsa, pp. 625–690.
- Kiessling, W., 2009. Geologic and biologic controls on the evolution of reefs. Annu. Rev. Ecol. Evol. Syst. 40, 173–192.
- Knoll, A.H., 2014. Paleobiological perspectives on early eukaryotic evolution. Cold Spring Harb. Perspect. Biol. 6, a016121.
- Knoll, A.H., Fischer, W.W., 2011. Skeletons and ocean chemistry: the long view. In: Gattuso, J.-P., Hansson, L. (Eds.), Ocean Acidification. Oxford University Press, Oxford, pp. 67–82.
- Laakso, T.A., Schrag, D.P., 2017. A theory of atmospheric oxygen. Geobiology 15, 366–384.
- Lau, K.V., Maher, K., Altiner, D., Kelley, B.M., Kump, L.R., Lehrmann, D.J., Silva-Tamayo, J.C., Weaver, K.L., Yu, M., Payne, J., 2016. Marine anoxia and delayed Earth system recovery after the end-Permian extinction. Proc. Natl. Acad. Sci. 113, 2360–2365.
- Lee, J.-H., Riding, R., 2018. Marine oxygenation, lithistid sponges, and the early history of Paleozoic skeletal reefs. Earth Sci. Rev. 181, 98–121.
- Leeder, M.R., 1982. Sedimentology: Process and Product. Chapman and Hall (344 pp). Lenton, T.M., Boyle, R.A., Poulton, S.W., Shields-Zhou, G.A., Butterfield, N.J., 2014. Coevolution of eukaryotes and ocean oxygenation in the Neoproterozoic era. Nat.
- Geosci. 7, 257–265. Lenton, T.M., Dahl, T.W., Daines, S.J., Mills, B.J.W., Ozaki, K., Saltzman, M.R., Porada, P., 2016. Earliest land plants created modern levels of atmospheric oxygen. Proc. Natl.
- Acad. Sci. 113, 9704–9709. Lerosey-Aubril, R., Paterson, J.R., Gibb, S., Chatterton, B.D.E., 2017. Exceptionally-pre-
- served late Cambrian fossils from the McKay Group (British Columbia, Canada) and the evolution of tagmosis in aglaspidid arthropods. Gondwana Res. 42, 264–279.
- Levin, L.A., 2003. Oxygen minimum zone benthos: adaptation and community response to hypoxia. Oceanogr. Mar. Biol. 41, 1–45.
- Levin, L.A., 2018. Manifestation, drivers, and emergence of open ocean deoxygenation. Annu. Rev. Mar. Sci. 10, 229–260.
- Levin, L.A., Ekau, W., Gooday, A.J., Jorissen, F., Middelburg, J.J., Naqvi, S.W.A., Neira, C., Rabalais, N.N., Zhang, J., 2009. Effects of natural and human-induced hypoxia on coastal benthos. Biogeosciences 6, 2063–2098.
- Lu, W., et al., 2018. Late inception of a resiliently oxygenated upper ocean. Science. https://doi.org/10.1126/science.aar5372.
- Lyons, T.W., Reinhard, C.T., Planavsky, N.J., 2014. The rise of oxygen in Earth's early ocean and atmosphere. Nature 506, 307–315.
- Martin, R.E., 1996. Secular increase in nutrient levels through the Phanerozoic: implications for productivity, biomass, and diversity of the marine biosphere. PALAIOS 11, 209–219.
- Matabos, M., Tunnicliffe, V., Juniper, S.K., Dean, C., 2012. A year in hypoxia: epibenthic community responses to severe oxygen deficit at a subsea observatory in a coastal inlet. PLoS One 7, e45626. https://doi.org/10.1371/journal.pone.0045626.
- Matysik, M., Al-Ramadan, K., Riding, R., 2015. Composition and morphology of early Devonian microbial and metazoan patch reefs: Qasr member of the Jauf Formation, northwestern Saudi Arabia. J. Sediment. Res. 85, 45–61.
- Mayhew, P.J., Jenkins, G.B., Benton, T.G., 2008. A long-term association between global temperature and biodiversity, origination and extinction in the fossil record. Proc. R. Soc. Lond. B Biol. Sci. 275, 47–53.
- Mayhew, P.J., Bell, M.A., Benton, T.G., McGowan, A.J., 2012. Biodiversity tracks temperature over time. Proc. Natl. Acad. Sci. 109, 15141–15145.
- McCook, L., Jompa, J., Diaz-Pulido, G., 2001. Competition between corals and algae on coral reefs: a review of evidence and mechanisms. Coral Reefs 19, 400–417.
- McKenzie, N.R., Hughes, N.C., Gill, B.C., Myrow, P.M., 2014. Plate tectonic influences on Neoproterozoic-early Paleozoic climate and animal evolution. Geology 42, 127–130. Meyer, K.M., Kump, L.R., 2008. Oceanic euxinia in Earth history: causes and con-
- sequences. Annu. Rev. Earth Planet. Sci. 36, 251–288.

- Mittelbach, G.G., Schemske, D.W., Cornell, H.V., Allen, A.P., Brown, J.M., Bush, M.B., Harrison, S.P., Hurlbert, A.H., Knowlton, N., Lessios, H.A., McCain, C.M., McCune, A.R., McDade, L.A., McPeek, M.A., Near, T.J., Price, T.D., Ricklefs, R.E., Roy, K., Sax, D.F., Schluter, D., Sobel, J.M., Turelli, M., 2007. Evolution and the latitudinal diversity gradient: speciation, extinction and biogeography. Ecol. Lett. 10, 315–331.
- Myshrall, K.L., Dupraz, C., Visscher, P.T., 2014. Patterns in microbialites throughout geologic time: is the present really the key to the past? In: Hembree, D.I., Platt, B.F., Smith, J.J. (Eds.), Experimental Approaches to Understanding. Topics in Geobiology. Springer, Dordrecht, pp. 111–142.
- Nursall, J.R., 1959. Oxygen as a prerequisite to the origin of the metazoa. Nature 183, 1170–1172.
- Och, L.M., Shields-Zhou, G.A., 2012. The Neoproterozoic oxygenation event: environmental perturbations and biogeochemical cycling. Earth Sci. Rev. 110, 26–57.
- Peters, S.E., Husson, J.M., Wilcots, J., 2017. The rise and fall of stromatolites in shallow marine environments. Geology 45, 487–490.
- Plummer, L.N., Sundquist, E.T., 1982. Total individual ion activity coefficients of calcium and carbonate in seawater at 25°C and 35%. Salinity, and implications to the agreement between apparent and thermodynamic constants of calcite and aragonite. Geochim. Cosmochim. Acta 46, 247–258.
- Pratt, B.R., 1982. Stromatolite decline-a reconsideration. Geology 10, 512-515.
- Pratt, B.R., 1995. The origin, biota and evolution of deep-water mud-mounds. In: Monty, C.L.V., Bosence, D.W.J., Bridges, P.H., Pratt, B.R. (Eds.), Carbonate Mud-Mounds: Their Origin and Evolution. International Association of Sedimentologists Special Publication. 28. Blackwell Science, pp. 49–123.
- Rabalais, N.N., Díaz, R.J., Levin, L.A., Turner, R.E., Gilbert, D., Zhang, J., 2010. Dynamics and distribution of natural and human-caused hypoxia. Biogeosciences 7, 585–619.
- Rasmussen, C.M.Ø., Ullmann, C.V., Jakobson, K.G., Lindskog, A., Hansen, J., Hansen, T., Eriksson, M.E., Dronov, A., Frei, R., Korte, C., Nielsen, A.T., Harper, D.A.T., 2016. Onset of main Phanerozoic marine radiation sparked by emerging Mid Ordovician icehouse. Sci. Rep. 18884.
- Reinhard, C.T., Planavsky, N.J., Olson, S.L., Lyons, T.W., Erwin, D.H., 2016. Earth's oxygen cycle and the evolution of animal life. Proc. Natl. Acad. Sci. 113, 8933–8938.
- Riding, R., 1992. Temporal variation in calcification in marine cyanobacteria. J. Geol. Soc. Lond. 149, 979–989.
- Riding, R., 1993. Phanerozoic patterns of marine CaCO₃ precipitation. Naturwissenschaften 80, 513–516.
- Riding, R., 2005. Phanerozoic reefal microbial carbonate abundance: comparisons with metazoan diversity, mass extinction events, and seawater saturation state. Rev. Esp. Micropaleontol. 37, 23–39.
- Riding, R., 2006. Microbial carbonate abundance compared with fluctuations in metazoan diversity over geological time. Sediment. Geol. 185, 229–238.
- Riding, R., Liang, L., 2005a. Geobiology of microbial carbonates: metazoan and seawater saturation state influences on secular trends during the Phanerozoic. Palaeogeogr. Palaeoclimatol. Palaeoecol. 219, 101–115.
- Riding, R., Liang, L., 2005b. Seawater chemistry control of marine limestone accumulation over the past 550 million years. Rev. Esp. Micropaleontol. 37, 1–11.
- Riding, R., Liang, L., Braga, J.C., 2014. Millennial-scale ocean acidification and late Quaternary decline of cryptic bacterial crusts in tropical reefs. Geobiology 12, 387–405.
- Royer, D.L., Berner, R.A., Montañez, I.P., Tabor, N.J., Beerling, D.J., 2004. CO₂ as a primary driver of Phanerozoic climate. GSA Today 14, 4–10.
- Royer, D.L., Donnadieu, Y., Park, J., Kowalczyk, J., Godde, Y., 2014. Error analysis of CO₂ and O₂ estimates from the long-term geochemical model GEOCARBSULF. Am. J. Sci. 314, 1259–1283.
- Saltzman, M.R., Young, S.A., Kump, L.R., Gill, B.C., Lyons, T.W., Runnegar, B., 2011. Pulse of atmospheric oxygen during the late Cambrian. Proc. Natl. Acad. Sci. 108, 3876–3881.
- Sandberg, P.A., 1975. New interpretations of Great Salt Lake ooids and of ancient nonskeletal carbonate mineralogy. Sedimentology 22, 497–537.
- Schneider, R.R., Schulz, H.D., Hensen, C., 2000. Marine carbonates: their formation and destruction. In: Schulz, H.D., Zabel, M. (Eds.), Marine Geochemistry. Springer, pp. 283–307.
- Schrag, D.P., Higgins, J.A., MacDonald, F.A., Johnston, D.T., 2013. Authigenic carbonate and the history of the global carbon cycle. Science 339, 540–543.
- Schubert, J.K., Bottjer, D.J., 1992. Early Triassic stromatolites as post-mass extinction disaster forms. Geology 20, 883–886.
- Schubert, J.K., Bottjer, D.J., 1995. Aftermath of the Permian-Triassic mass extinction event: paleoecology of Lower Triassic carbonates in the western USA. Palaeogeogr. Palaeoclimatol. Palaeoecol. 116, 1–39.
- Seibel, B.A., 2011. Critical oxygen levels and metabolic suppression in oceanic oxygen minimum zones. J. Exp. Biol. 204, 326–336.
- Sepkoski Jr., J.J., 1997. Biodiversity: past, present, and future. J. Paleontol. 71, 533-539.
- Servais, T., Lehnert, O., Li, J.U.N., Mullins, G.L., Munnecke, A., Nützel, A., Vecoli, M., 2008. The Ordovician biodiversification: revolution in the oceanic trophic chain.
- Lethaia 41, 99–109. Servais, T., Owen, A.W., Harper, D.A.T., Kröger, B., Munnecke, A., 2010. The Great
- Ordovician Biodiversification Event (GOBE): the palaeoecological dimension. Palaeogeogr. Palaeoclimatol. Palaeoecol. 294, 99–119. Servais, T., Perrier, V., Danelian, T., Klug, C., Martin, R., Munnecke, A., Nowak, H.,
- Nützel, A., Vandenbroucke, T.R.A., Williams, M., Rasmussen, C.M.Ø., 2016. The onset of the 'Ordovician Plankton Revolution' in the late Cambrian. Palaeogeogr. Palaeoclimatol. Palaeoecol. 458, 12–28.
- Sheehan, P.M., Harris, M.T., 2004. Microbialite resurgence after the late Ordovician extinction. Nature 430, 75–78.
- Soja, C.M., 1994. Significance of Silurian stromatolite-sphinctozoan reefs. Geology 22, 355–358.

Soja, C.M., Riding, R., 1993. Silurian microbial associations from the Alexander Terrane, Alaska. J. Paleontol. 67, 728–738.

- Sperling, E.A., Frieder, C.A., Raman, A.V., Girguis, P.R., Levin, L.A., Knoll, A.H., 2013. Oxygen, ecology, and the Cambrian radiation of animals. Proc. Natl. Acad. Sci. 110, 13446–13451.
- Stanley, S.M., 2007. An analysis of the history of marine animal diversity. Paleobiology 33, 1–55.
- Stramma, L., Johnson, G.C., Sprintall, J., Mohrholz, V., 2008. Expanding oxygenminimum zones in the tropical oceans. Science 320, 655–658.
- Stramma, L., Schmidtko, S., Levin, L.A., Johnson, G.C., 2010. Ocean oxygen minima expansions and their biological impacts. Deep-Sea Res. I Oceanogr. Res. Pap. 57, 587–595.
- Sun, X., Turchyn, A.V., 2014. Significant contribution of authigenic carbonate to marine carbon burial. Nat. Geosci. 7, 201–204.
- Svensson, J.R., Marshall, D.J., 2015. Limiting resources in sessile systems: food enhances diversity and growth of suspension feeders despite available space. Ecology 96, 819–827.
- Taylor, P.D., Wilson, M.A., 2003. Palaeoecology and evolution of marine hard substrate communities. Earth Sci. Rev. 62, 1–103.
- Trotter, J.A., Williams, I.S., Barnes, C.R., Lecuyer, C., Nicoll, R.S., 2008. Did cooling oceans trigger Ordovician biodiversification? Evidence from conodont thermometry. Science 321, 550–554.
- Tunnicliffe, V., 1981. High species diversity and abundance of the epibenthic community in an oxygen-deficient basin. Nature 294, 354–356.
- van de Schootbrugge, B., Wignall, P.B., 2015. A tale of two extinctions: converging end-Permian and end-Triassic scenarios. Geol. Mag. 153, 332–354.
- Van Roy, P., Orr, P.J., Botting, J.P., Muir, L.A., Vinther, J., Lefebvre, B., el Hariri, K., Briggs, D.E., 2010. Ordovician faunas of Burgess Shale type. Nature 465, 215–218. Vaquer-Sunyer, R., Duarte, C.M., 2008. Thresholds of hypoxia for marine biodiversity.

- Proc. Natl. Acad. Sci. 105, 15452-15457.
- Verberk, W.C., Bilton, D.T., Calosi, P., Spicer, J.I., 2011. Oxygen supply in aquatic ectotherms: partial pressure and solubility together explain biodiversity and size patterns. Ecology 92, 1565–1572.
- Wallace, M.W., Hood, A.vS., Shuster, A., Greig, A., Planavsky, N.J., Reed, C.P., 2017. Oxygenation history of the Neoproterozoic to early Phanerozoic and the rise of land plants. Earth Planet. Sci. Lett. 466, 12–19.
- Webb, G.E., 1996. Was Phanerozoic reef history controlled by the distribution of nonenzymatically secreted reef carbonates (microbial carbonate and biologically induced cement)? Sedimentology 43, 947–971.
- Webb, G.E., 2005. Quantitative analysis and paleoecology of earliest Mississippian microbial reefs, Gudman Formation, Queensland, Australia: not just post-disaster phenomena. J. Sediment. Res. 75, 877–896.
- Webby, B.D., 2002. Patterns of Ordovician reef development. In: Kiessling, W., Flügel, E., Golonka, J. (Eds.), Phanerozoic Reef Patterns. SEPM Special Publication 72. SEPM, Tulsa, pp. 129–179.
- Wilkinson, B.H., Walker, J.C.G., 1989. Phanerozoic cycling of sedimentary carbonate. Am. J. Sci. 289, 525–548.
- Wood, R., Erwin, D.H., 2018. Innovation not recovery: dynamic redox promotes metazoan radiations. Biol. Rev. 93, 863–873.
- Yao, L., Aretz, M., Chen, J., Webb, G.E., Wang, X., 2016. Global microbial carbonate proliferation after the end-Devonian mass extinction: mainly controlled by demise of skeletal bioconstructors. Sci. Rep. 6.
- Young, S.A., Gill, B.C., Edwards, C.T., Saltzman, M.R., Leslie, S.A., 2016. Middle–Late Ordovician (Darriwilian–Sandbian) decoupling of global sulfur and carbon cycles: Isotopic evidence from eastern and southern Laurentia. Palaeogeogr. Palaeoclimatol. Palaeoecol. 458, 118–132.
- Zaffos, A., Finnegan, S., Peters, S.E., 2017. Plate tectonic regulation of global marine animal diversity. Proc. Natl. Acad. Sci. 114, 5653–5658.